

# Periodic Oscillation of Gene Networks with Forcing and Delay

Yi Wang<sup>1,3</sup>      Zhongjun Ma<sup>2</sup>      Jianwei Shen<sup>1</sup>

<sup>1</sup>Institute of Systems Biology, Shanghai University, Shanghai, 200444, China;

<sup>2</sup>School of Mathematics and Computing Science,  
Guilin University of Electronic Technology, Guilin 541004, China;

<sup>3</sup>ZheJiang University of Finance and Economics, Hangzhou, 310012, China.

**Abstract** In this paper, we derive new criteria for evaluating the global stability of periodic oscillation for gene networks with small forcing and delay. Our results rely on the Lipschitz conditions of Hill function, topology of gene networks. In particular, Our method based on the proposed model transforms the original network into matrix analysis problem, thereby not only significantly reducing the computational complexity but also making analysis of periodic oscillation tractable for even large-scale nonlinear networks.

**Keywords** gene network; periodic oscillation

## 1 Introduction

It is well known that the gene networks (GNs) plays a key role in regulating dynamics of processes transducing genetic signals into phenotypic variation and integrating genomic information, environmental cues, and physiological or developmental stimuli. Thus, understanding the architectures and their design principles of GNs will fundamentally advance the study of core biological problems[1, 2]. In particular, since genetic regulatory networks are high-dimensional and nonlinear, it is indispensable to consider the network dynamics from the viewpoint of nonlinear system theory. However, how to appropriately represent real gene regulatory systems mathematically in terms of gene function, expression mechanisms, and signal-transduction pathways remains unclear. Mathematical models are useful for discovering higher order structure of an organism and for gaining deep insights into both static and dynamic behaviors of gene networks by extracting functional information from observation data [9].

A cellular system is generally characterized with significant time delays in gene regulation, in particular, for the transcription, translation, diffusion, and translocation processes. Moreover, periodic perturbations are widespread in the external environment(e.g. daily light-dark cycle and Moon's gravitational)and internal circumstance (e.g. cell division cycle or cellular motility). Such time delays and perturbation may affect the dynamics of the entire biological system, both qualitatively and quantitatively. Until now, most theoretical works on the study of coupling of gene oscillators[5, 6, 7, 8]. There have been some studies devoted to the stability and oscillations of GNs with fixed time-delay

[9, 10, 11]. It is shown that oscillations can be induced by delay in both nonstochastic and stochastic GNs.

Motivated by the above discussions, rather than rhythmic generators, the purpose of this paper are to study the periodic oscillation of GNs with small periodic forcing and time-delays. In particular, in this paper we derive new criteria for checking the global stability of periodic oscillation of GNs with SUM regulatory logic by using the continuation theorem of Mawhin's coincidence degree theory and Lyapunov functional. The proposed approach is general and can be applied to analyze many biological oscillations in an accurate manner. In other words, the theoretical results are able to cover a large range of nonlinear GNs even under uncertain environments. The paper is organized as follows. In Section 2, a framework of the general gene networks is given. In Section 3, we derive the main results and new criteria about stability of periodic oscillations with fixed time-delay. Section 4 provides an example to illustrate the application of these criteria. Several summary remarks are given in Section 5.

## 2 Model of gene network

The activity of a gene is regulated by other genes through the concentrations of their gene products, i.e. the transcription factors. Regulation can be quantified by the response characteristics, i.e. the level of gene expression as a function of the concentrations of transcription factors. In this paper, based on the structure of the gene network (GN) or the genetic regulatory network presented in [9], we consider a differential equation model described as follows [3, 9]:

$$\begin{aligned} \dot{m}_i(t) &= -a_i m_i(t) + b_i(p_1(t), \dots, p_n(t)), \\ \dot{p}_i(t) &= -c_i p_i(t) + d_i m_i(t), \quad i = 1, \dots, n. \end{aligned} \quad (1)$$

where  $m_i(t)$ ,  $p_i(t) \in \mathbb{R}$  are the concentrations of mRNA and protein of the  $i$ th node, respectively. In (1),  $a_i$  and  $c_i$  are the degradation rates of the mRNA and protein,  $d_i$  is synthesize rate of the protein, and  $b_i(t)$  is the regulatory function of the  $i$ th gene, which is a nonlinear function of the variables  $(p_1(t), \dots, p_n(t))$  but generally has a form of monotonicity with each variable [1, 4]. In this network, there is one output but multiple inputs for a single node or gene. A directed edge is linked from node  $j$  to  $i$  if the transcriptional factor or protein  $j$  regulates gene  $i$ .

Generally, the form of (1) may be very complicated, depending on all biochemical reactions involved in this regulation. Typical regulatory logics include AND-like gates and OR-like gates [19, 20] for  $b_i$ . In this paper, we focus on a model of gene networks where each transcription factor acts additively to regulate a gene. That is, the regulatory function is of the form  $b_i(p_1(t), \dots, p_n(t)) = \sum_{j=1}^n b_{ij}(p_j(t))$ , which is also called SUM logic [9, 21], i.e. the regulatory function sums over all the inputs. Such a SUM logic does exist in many natural genetic networks[19]. In synthetic gene networks, one of the simplest ways to implement such an additive input function is to provide a gene with multiple promoters, each responding to one of the inputs[10, 21]. Such a regulation by multiple promoters is indeed found in many gene systems.

The function  $b_{ij}(p_j(t))$  is generally expressed by a monotonic function of the Hill

form

$$b_{ij}(p_j(t)) = \begin{cases} e_{ij} \frac{(p_j(t)/k)^H}{1 + (p_j(t)/k)^H}, & \text{if transcription factor } j \text{ is an activator of gene } i; \\ e_{ij} \frac{1}{1 + (p_j(t)/k)^H}, & \text{if transcription factor } j \text{ is a repressor of gene } i. \end{cases} \quad (2)$$

where  $H$  is the Hill coefficient,  $k$  is a positive constant, and  $e_{ij} \geq 0$  is the dimensionless transcriptional rate of transcription factor  $j$  to gene  $i$ , which is a bounded constant.

If the time delay are introduced into system (1), (1) can be rewritten into the following equations:

$$\begin{aligned} \dot{m}_i(t) &= -a_i m_i(t) + \sum_{j=1}^n e_{ij} h_{ij}(p_j(t - \tau_j)) + \alpha_i(t), \\ \dot{p}_i(t) &= -c_i p_i(t) + d_i m_i(t - \sigma_i) + \beta_i(t), \end{aligned} \quad (3)$$

where  $\alpha_i(t) = \alpha_i(t + T)$  and  $\beta_i(t) = \beta_i(t + T)$  ( $i = 1, \dots, n$ ).

Throughout this paper, we always make the following assumption:

**A1.** There exists  $M_{ij} > 0$  such that  $|h_{ij}(u) - h_{ij}(v)| \leq M_{ij}|u - v|$  for each  $u, v \in \mathbb{R}$  ( $i, j = 1, \dots, n$ ).

Assumption **A1** is generally satisfied in GNs due to the saturation effects of transcription and translation processes. In order to obtain our main results, we need the following Lemma 1.

Assume that  $\mathbb{T}^{n \times n} = \{\mathbf{A} = (a_{ij})_{n \times n} : a_{ij} \leq 0, i \neq j\}$ .

**Lemma1.** ([24]): Let  $\mathbf{A} \in \mathbb{T}^{n \times n}$ . Then, each of the following conditions is equivalent to the statement ‘ $\mathbf{A}$  is a non-singular M-matrix’:

- (1) All of the principal minors of  $\mathbf{A}$  are positive.
- (2)  $\mathbf{A}$  has all positive diagonal elements and there exists a positive diagonal matrix  $\mathbf{D}$  such that  $\mathbf{AD}$  is strictly diagonally dominant, that is:

$$a_{ii}d_i > \sum_{i \neq j} |a_{ij}|d_i, \quad i = 1, 2, \dots, n.$$

- (3)  $\mathbf{A}$  is inverse-positive, that is,  $\mathbf{A}^{-1}$  exists and  $\mathbf{A}^{-1} \geq 0$ .

The above Lemma 1 is widely used in the theoretical analysis of optimization design, finance mathematics, and neural networks. In this paper, we adopt Lemma 1 to estimate the norm of  $m_i(t)$  and  $p_i(t)$  in the main results, i.e. Theorem I of next section.

### 3 Some results of stability in Gene Networks with fixed delay

In this section, we derive the main results which ensure the existence and stability of periodic oscillators in GNs with forcings and delay, where forcing  $\alpha_i(t)$  and  $\beta_i(t)$  are assumed to be small and periodic. Letting  $\alpha_i^+ = \sup_{t \geq 0} |\alpha_i(t)|$  and  $\beta_i^+ = \sup_{t \geq 0} |\beta_i(t)|$ , we have the following sufficient conditions on the existence of periodic solutions:

**Theorem I.** Assume that  $\alpha_i^+$  and  $\beta_i^+$  are bounded number, (A1) hold. Then, the system (3) has at least one  $T$ -periodic solution corresponding to the same periodic of forcing, if

$$\Gamma = \begin{pmatrix} I_n & \Gamma_{12} \\ \Gamma_{21} & I_n \end{pmatrix}$$

is a non-singular M-matrix, where  $I_n$  is a unit matrix and  $\Gamma_{12} = (u_{ij})_{n \times n}$ ,  $u_{ij} = -e_{ij}M_{ij}/a_i$ ;  $\Gamma_{21} = \text{diag}\{-d_1/c_1, -d_2/c_2, \dots, -d_n/c_n\}$ .

The detail proof of this result is given in [30]. To ensure non-singular M-matrix, all of the elements  $\Gamma_{12}$  and  $\Gamma_{21}$  should be relatively small, comparing with the diagonal elements. In other words,  $a_i$  and  $c_i$  are required to be relatively large, comparing with the synthetic rates. With those conditions, (3) is ensured to have one T-periodic solution.

In following theorem, we show that under certain conditions system (3) has at least one T-periodic solution which is globally attractive. It means that all the trajectories of (3) eventually converge to the unique periodic solution.

**Theorem II.** Assume that all conditions of Theorem I hold. The system (3) has a unique T- periodic solution, which is globally attractive, if

$$\mathbb{O} = \begin{pmatrix} \mathbb{O}_{11} & \mathbb{O}_{12} \\ \mathbb{O}_{21} & \mathbb{O}_{22} \end{pmatrix}$$

is a non-singular M-matrix, where  $\mathbb{O}_{11} = \text{diag}(2a_1 - \sum_{j=1}^n e_{1j}M_{1j}, 2a_2 - \sum_{j=1}^n e_{2j}M_{2j}, \dots, 2a_n - \sum_{j=1}^n e_{nj}M_{nj})$ ;  $\mathbb{O}_{12} = (v_{ij})_{n \times n}$ ,  $v_{ij} = -e_{ij}M_{ij}$ ;  $\mathbb{O}_{21} = \text{diag}(-d_1, -d_2, \dots, -d_n)$ ;  $\mathbb{O}_{22} = \text{diag}(2c_1 - d_1, 2c_2 - d_2, \dots, 2c_n - d_n)$ .

The proof is given in [30]. The conditions for Theorem II are similar to those of Theorem I, i.e. the degradation rates are required to be larger than those of the synthesis rates so that  $\mathbb{O}$  is a non-singular M-matrix. Next, we further give the conditions for the asymptotical stability of the T-periodic solution.

**Theorem III.** Assume that all conditions of Theorem I hold. The system (3) has a unique T- periodic solution, which is globally and asymptotically stable if  $\mathbb{O} \in \mathbb{T}$  and is a weakly column diagonally dominant matrices.

The detail proof is given in [30]. The conditions of Theorem III also require large degradation rates, comparing with the synthesis rates. As indicated by the theoretical results, a network with forcing (even with sufficiently small forcing) has globally stable oscillation under certain conditions. It means that all the trajectories of (3) will eventually reach the unique periodic solution. It's worth noting that, even if the parameters and regulatory functions are uncertain, we may design a robust GN, which can be driven by any periodic signals and thereby is able to be synchronized with the driven forces even under uncertain environment, as long as the parameter intervals are finite.

In this section, we present an example of a gene network to show the effectiveness and correctness of our theoretical results.

We consider the dynamics of the repressilator, which has been theoretically predicted and experimentally investigated in *Escherichia coli* [3]. The repressilator is a cyclic negative-feedback loop comprising three repressor genes (*lacl*, *tetR* and *cl*) and their promoters. The kinetics of the system are determined by six coupled first-order differential equations

$$\begin{aligned} \dot{m}_i(t) &= -a_i m_i(t) + \frac{e}{1 + p_j(t - \tau_i)^2} + \alpha_i(t), \\ \dot{p}_j(t) &= -c_j p_j(t) + d_j m_j(t - \sigma_j) + \beta_j(t), \\ i &= lacl, tetR, cl; j = cl, lacl, tetR, \end{aligned} \quad (4)$$

where  $m_i$  and  $p_i$  are the concentrations of the three mRNAs and repressor-proteins, and  $e$  denotes the ratio of the protein decay rate to the mRNA decay rate. We select a set of

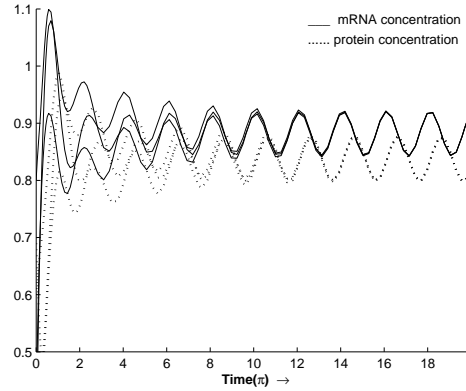


Figure 1: Time evolution of three mRNA and protein concentrations of GN (4)

biologically plausible parameters as  $e = 1.5$ ,  $a_i = c_i = 1$ ,  $d_i = 0.95$ ,  $\tau_i = 0.4$ ,  $\sigma_j = 0.6$ , and choose small periodic perturbations as  $\alpha_i(t) = 0.025 \cos t$ ,  $\beta_i(t) = 0.025 \sin t$  with  $T = 2\pi$ , which are assumed to be weakly coupled from external environment or other rhythmic generators.

By appropriate computation, we have  $\max h' = \frac{3\sqrt{3}}{8} < 21/32$ ,

$$\Gamma = \begin{pmatrix} 1 & 0 & 0 & 0 & 0 & -\frac{63}{64} \\ 0 & 1 & 0 & -\frac{63}{64} & 0 & 0 \\ 0 & 0 & 1 & 0 & -\frac{63}{64} & 0 \\ -\frac{19}{20} & 0 & 0 & 1 & 0 & 0 \\ 0 & -\frac{19}{20} & 0 & 0 & 1 & 0 \\ 0 & 0 & -\frac{19}{20} & 0 & 0 & 1 \end{pmatrix}$$

and

$$\mathbb{O} = \begin{pmatrix} \frac{65}{64} & 0 & 0 & 0 & 0 & -\frac{63}{64} \\ 0 & \frac{65}{64} & 0 & -\frac{63}{64} & 0 & 0 \\ 0 & 0 & \frac{65}{64} & 0 & -\frac{63}{64} & 0 \\ -\frac{19}{20} & 0 & 0 & \frac{21}{20} & 0 & 0 \\ 0 & -\frac{19}{20} & 0 & 0 & \frac{21}{20} & 0 \\ 0 & 0 & -\frac{19}{20} & 0 & 0 & \frac{21}{20} \end{pmatrix}.$$

It is easy to check that  $\Gamma$  and  $\mathbb{O}$  are non-singular M-matrices. From Theorem 3, the

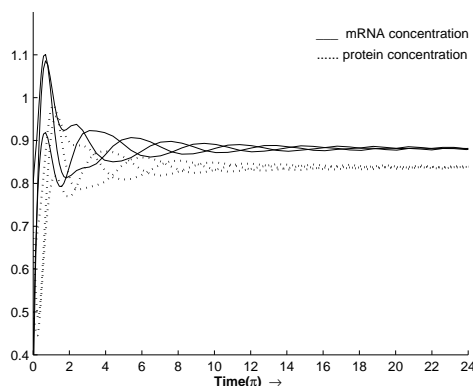


Figure 2: Time evolution of three mRNA and protein concentrations of GN(4) without forcing.

repressilator with this set of parameters has a  $T$ -Periodic oscillation, which is globally and asymptotically stable. In Fig.1 we plot the trajectories of mRNAs and proteins concentrations, which confirm the theoretical prediction.

From other study[5, 9, 12], we knew delay always induce the periodic oscillation. Here, by numerical simulation, we find that GN have only stable stationary solution absent small forcing(i.e.,  $\alpha_i(t) = \beta_i(t) = 0$ ), see Fig.2. It is sufficiently demonstrate that the small extra forcing can produce periodic oscillation.

## 4 Numerical Example

In this section, we present an example of a gene network to show the effectiveness and correctness of our theoretical results.

We consider the dynamics of the repressilator, which has been theoretically predicted and experimentally investigated in *Escherichia coli* [3]. The repressilator is a cyclic negative-feedback loop comprising three repressor genes (*lacl*, *tetR* and *cl*) and their promoters. The kinetics of the system are determined by six coupled first-order differential equations

$$\begin{aligned} \dot{m}_i(t) &= -a_i m_i(t) + \frac{e}{1 + p_j(t - \tau_i)^2} + \alpha_i(t), \\ \dot{p}_j(t) &= -c_j p_j(t) + d_j m_j(t - \sigma_j) + \beta_j(t), \\ i &= lacl, tetR, cl; j = cl, lacl, tetR, \end{aligned} \quad (5)$$

where  $m_i$  and  $p_i$  are the concentrations of the three mRNAs and repressor-proteins, and  $e$  denotes the ratio of the protein decay rate to the mRNA decay rate. We select a set of biologically plausible parameters as  $e = 1.5$ ,  $a_i = c_i = 1$ ,  $d_i = 0.95$ ,  $\tau_i = 0.4$ ,  $\sigma_j = 0.6$ , and choose small periodic perturbations as  $\alpha_i(t) = 0.025 \cos t$ ,  $\beta_i(t) = 0.025 \sin t$  with  $T = 2\pi$ , which are assumed to be weakly coupled from external environment or other rhythmic generators.

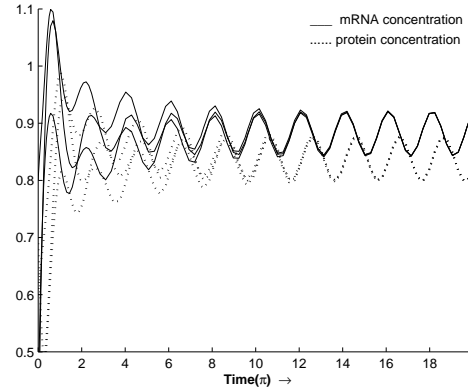


Figure 3: Time evolution of three mRNA and protein concentrations of GN (4)

By appropriate computation, we have  $\max h' = \frac{3\sqrt{3}}{8} < 21/32$ ,

$$\Gamma = \begin{pmatrix} 1 & 0 & 0 & 0 & 0 & -\frac{63}{64} \\ 0 & 1 & 0 & -\frac{63}{64} & 0 & 0 \\ 0 & 0 & 1 & 0 & -\frac{63}{64} & 0 \\ -\frac{19}{20} & 0 & 0 & 1 & 0 & 0 \\ 0 & -\frac{19}{20} & 0 & 0 & 1 & 0 \\ 0 & 0 & -\frac{19}{20} & 0 & 0 & 1 \end{pmatrix}$$

and

$$\mathbb{O} = \begin{pmatrix} \frac{65}{64} & 0 & 0 & 0 & 0 & -\frac{63}{64} \\ 0 & \frac{65}{64} & 0 & -\frac{63}{64} & 0 & 0 \\ 0 & 0 & \frac{65}{64} & 0 & -\frac{63}{64} & 0 \\ -\frac{19}{20} & 0 & 0 & \frac{21}{20} & 0 & 0 \\ 0 & -\frac{19}{20} & 0 & 0 & \frac{21}{20} & 0 \\ 0 & 0 & -\frac{19}{20} & 0 & 0 & \frac{21}{20} \end{pmatrix}.$$

It is easy to check that  $\Gamma$  and  $\mathbb{O}$  are non-singular M-matrices. From Theorem 3, the repressilator with this set of parameters has a  $T$ -Periodic oscillation, which is globally and asymptotically stable. In Fig.1 we plot the trajectories of mRNAs and proteins concentrations, which confirm the theoretical prediction.

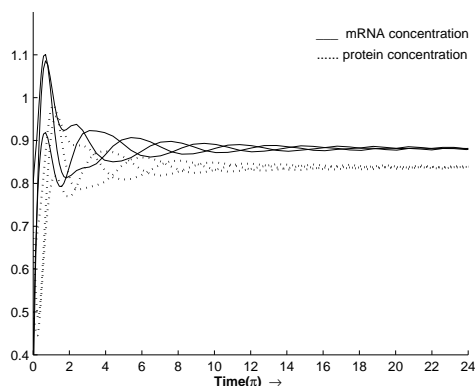


Figure 4: Time evolution of three mRNA and protein concentrations of GN(4) without forcing.

From other study[5, 9, 12], we knew delay always induce the periodic oscillation. Here, by numerical simulation, we find that GN have only stable stationary solution absent small forcing(i.e.,  $\alpha_i(t) = \beta_i(t) = 0$ ), see Fig.2. It is sufficiently demonstrate that the small extra forcing can produce periodic oscillation.

## 5 Discussion.

Generally speaking, a cellular system is not only affected by various external fluctuations but also characterized with significant time delays in gene regulation, in particular, for the transcription, translation, diffusion, and translocation processes. Such time delays may affect the dynamics of the entire biological system both qualitatively and quantitatively. In this paper, we derived new criteria for checking the global stability of periodic oscillation of GNs with delays and periodic forcings by using the continuation theorem of Mawhin's coincidence degree theory, the non-singular M-matrix and Lyapunov function. Theoretical results ensure that the GN has a stable periodic oscillation provided that there is a small periodic driving force, which can be used to analyze cellular oscillations, to understand synchronization phenomena, and even to design a robust synthetic oscillator.

### 5.1 Designing a large scale GN

As indicated in this paper, the networks with forcing have stable oscillation under certain conditions which rely on the Lyapunov constants of the regulatory functions, dimensionless transcriptional rate, and the degradation rates of mRNAs or proteins. When analyzing or designing a large scale GN, we need the following steps based on Theorems I-III.

- 1) Firstly, we select a set of biologically plausible parameters and compute the Lipschitz constants  $M_{ij}$  of the function  $b_i$ .
- 2) Check whether  $\Gamma$  and  $\mathbb{O}$  are non-singular M-matrices.

If all of the conditions are satisfied, there exists a stable periodic oscillation, which



coincides with the weak periodic signal in phase. Owing to the fact that our results are robust for genetic oscillator networks with forcing and distributed delay, we can design a robust oscillation in the GN within the defined parameter intervals or ranges.

## 5.2 Small perturbation can also induce oscillation

Since genetic regulatory networks are high-dimensional and nonlinear, it is indispensable to consider the network dynamics from the viewpoint of nonlinear systems theory. Up to now, several theoretical models have been successfully developed to understand circadian phenomena of GNs, but few studies consider the small perturbations. On the other hand, these perturbations always exist in the GNs, such as the stochastic regulation, production and decay processes which are due to temporary chemical or physical changes in the environment, or periodic perturbations from other cells. Unfortunately, previous results are mainly for the perturbations on linear systems, and there are few results on the perturbations of nonlinear systems due to the mathematical difficulties.

In [23], a model for a synthetic gene oscillator is present, and the coupling of the small oscillator to a periodic process that is intrinsic to the cell is considered. They studied the synchronization properties of the coupled system, and showed how the oscillator can be constructed to yield a significant amplification of cellular oscillations both numerically and experimentally.

In our paper, we consider a general nonlinear GN model and show that under certain conditions, the nonlinear system coupled with even small perturbation can always oscillate with the signals.

## Acknowledges

This work is supported by the NNSF of China: No. 10672093. The authors thank Prof. Zengrong Liu and Prof. Luonan Chen for helpful discussions and suggestions.

## References

- [1] Smolen, P., Baxter, D.A. and Byrne, J. H., Mathematical modeling of gene networks, *Neuron*, vol. 26(3) (2000)567-580.
- [2] Smolen, P., Baxter, D.A., and Byrne, J.H., Modeling transcriptional control in Gene Networks—methods, recent results, and future directions, *Bulletin of Mathematical Biology*, 62(2000) 247–292
- [3] Elowitz, M.B. and Leibler, S., A synthetic oscillatory network of transcriptional regulators, *Nature*,403( 2000)335-338.
- [4] Gardner, T.S., Cantor, C.R. and Collins, J.J., Construction of a genetic toggle switch in *Escherichia Coli*, *Nature*, 403(2000)339-342.
- [5] McMillen, D., Kopell, N., Hasty, J., and Collins, J., Synchronizing genetic relaxation oscillators by intercell signaling, *Natl. Acad. Sci., U.S.A.*, 99(2002)679-684.
- [6] Wang, R., Chen, L., Synchronizing genetic oscillators by signaling molecules. *J Biol Rhythms*, 20(2005)257-269.
- [7] Li, C., Chen, L., and Aihara, K., synchronization of coupled nonidentical genetic oscillators, *Phys. Bio.*3(2006)37-44.

- [8] Li, C., Chen, L. and Aihara, K., Stochastic synchronization of genetic oscillator networks, *BMC Syst. Biol.*, 1(2007), DOI:10.1186/1752-0509-1-6.
- [9] Chen, L., Aihara, K., Stability of genetic regulatory networks with time delay. *IEEE Trans. Circuits Syst. I.* 49(2002)602-608.
- [10] Li, C., Chen, L., Stability of Genetic Networks With SUM Regulatory Logic, Lur'e System and LMI Approach, *IEEE Trans. Circuits Syst. I.* 53(2006)2451-2458.
- [11] Bratsun, D., Volfson, D., Tsimring, L.S., and Hasty, J., Delay-induced stochastic oscillations in gene regulation", *Natl. Acad. Sci., U.S.A.*, (2005)14593-14598.
- [12] Rateitschak, K. and Wolkenhauer, O., Intracellular delay limits cyclic changes in gene expression, *Mathematical Biosciences*, 205, (2007)163-179.
- [13] Elowitz, M.B., Levine, A.J., Siggia, E.D., Swain PS, Stochastic gene expression in a single cell. *Science*, 297(2002)1183-1186.
- [14] Paulsson, J., Summing up the noise in gene networks. *Nature*, 427(2004)415-418.
- [15] Raser, J.M., O'Shea, E.K., Noise in gene expression, Origins, consequences, and control. *Science*, 309(2005)2010-2013.
- [16] Blake, W.J., Kaern, M., Cantor, C.R., Collins J.J., Noise in eukaryotic gene expression. *Nature*, 422(2003)633-637.
- [17] Kaern, M., Elston, T.C., Blake, W.J. and Collins, J.J., Stochasticity in gene expression: From theories to phenotypes. *Nature Reviews Genetics*, 6(2005)451-464.
- [18] Li, C., Chen, L., Aihara, K., Transient resetting: A novel mechanism for synchrony and its biological examples. *PLoS Comp Biol* 2(2006)e103.
- [19] Yuh, C.H., Bolouri, H., and Davidson, E.H., Genomic cis-regulatory logic, Experimental and computational analysis of a sea urchin gene, *Science*, 279(1998)1896-1902.
- [20] Buchler, N.E., Gerland, U. and Hwa, T., On schemes of combinatorial transcription logic, *Proc. Natl. Acad. Sci. USA*, 100(2003)5136-5141.
- [21] Kalir, S., Mangan, S. and Alon, U., A coherent feed-forward loop with a SUM input function prolongs flagella expression in *Escherichia coli*, *Molecular Syst. Biol.*(2005) 10.1038/msb4100010.
- [22] Hale, J.K., Verduyn Lunel, S.M., introduction to Functional Differential Equations, fourth ed., Springer, New York, 1993.
- [23] Hasty, J., Dolnik, N., Rottschaefer, V. and Collins, J.J., Synthetic Gene Network for Entraining and Amplifying Cellular Oscillations, *PRL.*, V88, 14 (2002)148101.
- [24] Berman, A and Plemmons, R. J., *Nonnegative Matrices in The Mathematical Science*, 1979 (New York, Academic).
- [25] Dunlap, J.C., Molecular bases for circadian clocks. *Cell* 96(1999) 271-290.
- [26] Yisraeli, J.K., Sokol, S. and Melton, D.A., A two-step model for the localization of maternal mRNA in *Xenopus* oocytes, involvement of microtubules and microfilaments in the translocation and anchoring of Vg1 mRNA, *Development* 108(1990) 289-298.
- [27] King, M.L., Molecular basis for cytoplasmic localization. *Dev. Genet.* 19(1996)183-189.

- [28] Sabry, J., O'Connor, T. and Kirschner, M. W., Axonal transport of tubulin in T11 Pioneer neurons in situ. *Neuron* 14( 1995)1247-1256.
- [29] Jacobs, C. and L. Shapiro, Microbial asymmetric cell division, localization of cell fate determinants. *Curr. Opin. Gen. Dev.* 8, (1998)386-391.
- [30] Wang, Y., Ma, Z.J., Shen, J.W., Liu, Z.R., Chen, L.N., Periodic oscillation of gene networks with small forcing and distributed delay, submitting.