

Antitumor mechanism research of cryptotanshinone by module-based network analysis

Shichao Zheng, Zhenzhen Ren, Shifeng Wang, Yanling Zhang*, Yanjiang Qiao*

School of Chinese Pharmacy
Beijing University of Chinese Medicine
Beijing, 100102, China
zsc305@hotmail.com

Abstract—Cryptotanshinone (CPT) is one of the major liposoluble ingredients in *Salvia miltiorrhiza* which exerts antitumor activity on several types of cancers. However, the action mechanism of CPT remained to be clarified. The current study aimed to elucidate the antitumor mechanism of CPT based on the protein interaction network (PIN) analysis. A PIN of CPT was constructed with 244 nodes and 778 interactions, and was analyzed by Gene ontology (GO) enrichment analysis based on Markov Cluster algorithm (MCL). Two modules were found to be intimately associated with antitumor. Still further, the antitumor effect of CPT may be partly attributable to inhibiting the activation of the c-Src pathway and overexpression of EGFR, to mediating overexpression of PIAS and activation of EIF2AK3. Therefore, this study may shed new light on the antitumor mechanism and treatment of CPT.

Keywords—Protein interaction network; module; antitumor actions; cryptotanshinone; GO enrichment analysis

I. INTRODUCTION

Salvia miltiorrhiza (Danshen) is a classical traditional Chinese herbal medicine (TCM) with approximately 1,000 years of clinical application for the treatment of various kinds of diseases [1–4]. In the Dictionary of Traditional Chinese Medicine Prescription, Danshen ranks No.4 among the 1362 classic and empirical prescriptions for cancer therapy by frequency analysis [5]. Therefore, Danshen plays an important role in TCM for cancer treatment. Cryptotanshinone (CPT), a major active component isolated from *Salvia miltiorrhiza*, has been shown to possess various pharmacological activities, such as antitumor, anti-inflammatory and antibacterial [6–8]. Antitumor activity of CPT had become a research focus in recent years. However, the antitumor mechanism of CPT was not fully understood.

Network pharmacology is a novel subject to discover TCM from a systems perspective and at the molecular level [9]. Proteins are vital macromolecules, at both cellular and systemic levels, but they rarely act alone. Protein-protein interactions (PPIs) are major bearers of the biological process. The relevance of PPI as putative therapeutic targets for the

development of new treatment is particularly evident in cancer, with several ongoing clinical trials within this area [10]. The GO[11] project is a collaborative effort to construct ontologies which facilitate biologically meaningful annotation of gene products. It provides a collection of well-defined biological terms, spanning biological processes, molecular functions and cellular components. GO enrichment is a common statistical method used to identify shared associations between proteins and annotations to GO. Module-network and GO analysis may provide an efficient way to illustrate the molecular mechanism of antitumor action for CPT.

In this study, a network pharmacology approach was applied to analyze the antitumor mechanisms of CPT. PPIs were adopted in constructing a biological network. And the characteristics of scale-free, small-world network and module were analyzed. This paper aimed to further elucidate the antitumor molecular mechanism of CPT, and provide reference for its clinical application and further drug development.

II. MATERIALS AND METHODS

A. Network construction

The targets information of CPT was extracted from ChEMBL (<https://www.ebi.ac.uk/chembl/#>) and STITCH 3.1 (<http://stitch.embl.de/>). ChEMBL is an open large-scale bioactivity database, with the information largely manually extracted from the medicinal chemistry literatures [12]. STITCH [13] is a database for protein–chemical interactions that integrates many sources of experimental and manually curated evidence with text-mining information and interaction predictions. Every protein–chemical interaction has a confidence score.

The PPIs information was obtained from the online update databases of String 9.1 (<http://string-db.org>) which provides uniquely comprehensive coverage and ease of access to both experimental as well as predicted PPI information. Thereafter, the known and predicted associations are scored and integrated [14].

This work was supported by the National Key Technology R&D Program (No.2008BAI51B01) and the National Natural Science Foundation of China (No.81173522). *Correspondence author: Y.Zhang(colleen_zhang@163.com) Y.Qiao (yjqiao@263.net).

B. Network analysis

The topological properties of the PPI network, such as degree distribution, clustering coefficient and average shortest path were analyzed by Network Analyzer [15] in Cytoscape software. Degree distribution refers to the number of connections between proteins of the network; Clustering

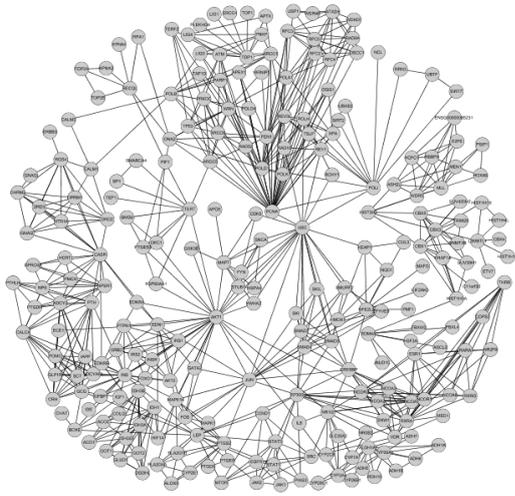


Fig. 1. Protein interaction network of CPT. The nodes and edges indicated the proteins and their relations

coefficient is the average density of the node neighborhoods; Average shortest path means the average density of shortest paths between all pairs of nodes [16]. Properties of scale-free, small world and modularity of the PIN were also investigated based on the topological parameters.

Functional modules of the network were explored by the MCL [17]. MCL simulated a flow on the graph by calculating successive powers of the associated adjacency matrix and the value of the inflation parameter strongly influenced the number of clusters, which was superior to other algorithms, e.g. RNSC, MCODE and SPC with highlighting the robustness to graph alterations [18]. Based on the identified modules, GO enrichment analysis was utilized to predict possible biological roles of the modules by evaluating the involved biological processes, using the BinGO [19] plugin for Cytoscape.

III. RESULTS AND DISCUSSION

A. Construction of the network

6 and 30 human proteins were extracted from STITCH 3.1 and ChEMBL (up to January of 2014), respectively. 35 human proteins were obtained as CPT targets after removing a repeat protein. Research had shown that the binding affinity (IC_{50}) of CPT and STAT3 was 4600nm, however, the IC_{50} are not available because CPT can inhibit or activate other proteins [20, 21]. The targets were listed in Table 1. PPIs information of targets whose confidence score was higher than 0.7 was imported in Cytoscape 2.8.3 [22], then carried union calculation and removed duplicated edges of PPIs using Advanced Network Merge [23] of Plugins, lastly, selected the

largest connected subgraph as the PIN of CPT which included 244 nodes and 778 edges. The PIN of CPT was shown in figure 1.

B. Network analysis

1) Topological analysis

TABLE I. THE LIST OF TARGETS OF CPT.

Targets	UniProt ID	Source	Targets	UniProt ID	Source
ALDH1A1	P00352	ChEMBL	POLH	Q9Y253	ChEMBL
ATAD5	Q96QE3	ChEMBL	POLI	Q9UNA4	ChEMBL
BAZ2B	Q9UIF8	ChEMBL	POLK	Q9UBT6	ChEMBL
CBX1	P83916	ChEMBL	RECQL	P46063	ChEMBL
FEN1	P39748	ChEMBL	RGS4	P49798	ChEMBL
GLP1R	P43220	ChEMBL	SMAD3	P84022	ChEMBL
GMNN	O75496	ChEMBL	STAT3	P40763	BOTH
IDH1	O75874	ChEMBL	TDP1	Q9NUW8	ChEMBL
KAT2A	Q92830	ChEMBL	TERT	O14746	ChEMBL
KDM4A	O75164	ChEMBL	THRB	P10828	ChEMBL
KDM4E	B2RXH2	ChEMBL	VDR	P11473	ChEMBL
L3MBTL1	Q9Y468	ChEMBL	WRN	Q14191	ChEMBL
MAPT	P10636	ChEMBL	BCHE	P06276	STITCH
MBNL1	Q9NR56	ChEMBL	INS	P01308	STITCH
MLL	Q03164	ChEMBL	EDN1	P05305	STITCH
NFE2L2	Q16236	ChEMBL	PTGS2	P35354	STITCH
NPSR1	Q6W5P4	ChEMBL	NR1I2	O75469	STITCH
POLB	P06746	ChEMBL			

All the topological parameters were calculated and shown in Table 2.

As shown in Figure 2A, the degree distribution of the PIN of CPT followed the power law distribution and the equation is $y=85.694x^{-1.181}$. So the PIN of CPT was a scale-free network that possessed fragility and robustness [24-26].

As shown in Figure 2B, network path length was mostly concentrated in 3-5 steps. The shortest path length between any two proteins of 4.958 links was calculated. Small world networks have a property that Characteristic path length is small [27]. This indicated that most proteins were closely

TABLE II. THE SIMPLE PARAMETERS OF PROTEIN INTERACTION NETWORK OF CPT

Parameters	PIN of CPT
Clustering coefficient	0.659
Connected components ^a	1
Network diameter ^b	12
Network centralization ^c	0.123
Shortest path	59292(100%)
Characteristic path length	4.958

Network heterogeneity ^d	0.743
------------------------------------	-------

^a. The connected component is 1 that indicates the network has no other subgraph.

^b. The network diameter is the greatest distance between any pair of vertices

^c. Network centralization is a network index that measures the degree of dispersion of all node centrality scores in a network

^d. network heterogeneity can characterize the degree of uneven distribution of the network

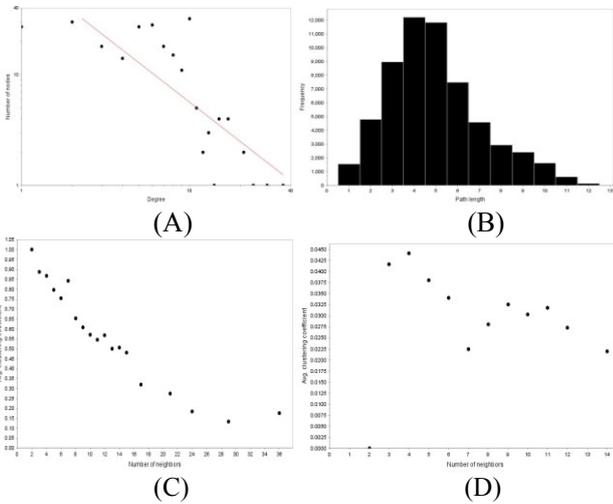


Fig. 2. Topological properties of network. (A) The degree distribution of CPT network; (B) Shortest path length distribution of CPT network; (C) Average clustering coefficient of CPT network; (D) Average clustering coefficient of random network

linked and the PIN of CPT was a small world network.

As shown in Figure 2 (C and D), compared with random network whose number of nodes and edges were identical to the PIN of CPT, the clustering coefficient of PIN was higher. In graph theory, clustering coefficient is the measure of the clustering degree of nodes. It indicated the PIN of CPT was more modularity. These results suggested that the network exhibited scale-free property, small world property and modular architecture.

2) Clustering and GO enrichment analysis

22 modules were identified with MCL algorithm (shown in Figure 3). All 22 modules included 236 of the total 244 proteins.

The results of functional enrichment analysis using BinGO are shown in Table 3, showed that CPT played a pharmacodynamics with the biological processes, such as DNA metabolic process, tricarboxylic acid metabolic process, icosanoid metabolic process, antitumor response, etc. And module 8 and 13 are related to antitumor action.

Module 8 contained proteins such as STAT3, JAK1, JAK2, PIAS3, SRC, EGFR. As a member of STAT family which transduces extracellular signals and regulates transcription of target genes, STAT3 has been proved to be the most intimately linked to tumorigenesis [28]. Dae-Seop Shin had studied that CPT exerted antitumor property by inhibiting STAT3 [20]. STAT3 is activated by JAK [21] and dysregulation of the pathway is frequently observed in primary tumors [29]. It was previously reported that CPT

induced inhibition of breast tumor growth through the JAK/STAT signaling pathway [30]. PIAS proteins were important transcriptional co-regulators of the JAK/STAT growth when overexpressed [31]. CPT may possess antitumor activity associating with overexpression of PIAS. SRC and STAT3 are coordinately altered in many human signaling pathway and PIAS inhibited human lung cancer cell tumors [32-34]. The activation of the c-Src pathway leading to the promotion of survival, angiogenesis, proliferation and invasion pathways has been observed in about 50% of tumors [35]. SRC was activated by EGFR [36] while EGFR overexpression or overactivation had been associated with a number of cancers, including lung cancer, anal cancers, which produced uncontrolled cell division [37]. The analysis of module 8 indicated that the antitumor effects of CPT may be attributed to inhibiting the activation of the c-Src pathway and overexpression of EGFR.

Antitumor activity of Module 13 was closely related to regulation of apoptotic process including HMOX1, EIF2AK3, etc. HMOX1 belongs to the heme oxygenase family, which was activated at high concentrations of heme and was thought to function as an oxidative stress indicator [38]. It was reported that HMOX1 was involved in estrogen-induced cell apoptosis progress. Antitumor effects of CPT may be relevant to HMOX1. EIF2AK3, also known as PERK, is a type I membrane protein located in the endoplasmic reticulum (ER) where it was induced by ER stress [39]. However, activation of PERK resulted in phosphorylation of eukaryotic translation initiation factor 2 subunit α (eIF2 α), which lead to

TABLE III. GO BIOLOGICAL PROCESS TERMS OF THE MODULES DISPLAY PARTIALLY

Modules	GO terms	P-value
Module 1	cellular response to endogenous stimulus	7.28E-36
Module 2	response to DNA damage stimulus	1.89E-39
Module 3	transcription initiation from RNA polymerase II promoter	4.95E-25
Module 4	xenobiotic metabolic process	8.62E-21
Module 5	histone lysine methylation	1.41E-12
Module 6	tricarboxylic acid cycle	1.57E-18
Module 7	chromatin organization	2.84E-10
Module 8	JAK-STAT cascade	1.71E-09
Module 9	telomere maintenance	6.97E-12
Module 10	icosanoid biosynthetic process	1.74E-15
Module 11	adenylate cyclase-modulating G-protein coupled receptor signaling pathway	5.42E-13
Module 12	double-strand break repair	1.10E-10
Module 13	regulation of apoptotic process	3.59E-05
Module 14	transcription initiation from RNA polymerase I promoter	1.25E-05
Module 15	mitotic recombination	3.52E-11
Module 16	chromatin organization	2.04E-04
Module 17	chromatin organization	3.07E-08
Module 18	neuropeptide signaling pathway	1.03E-06

Module 19	neural crest cell development	2.44E-10
Module 20	synaptic transmission, cholinergic	1.36E-06
Module 21	steroid metabolic process	1.11E-06

Module 22	regulation of cell communication by electrical coupling involved in cardiac conduction	1.94E-08
-----------	--	----------

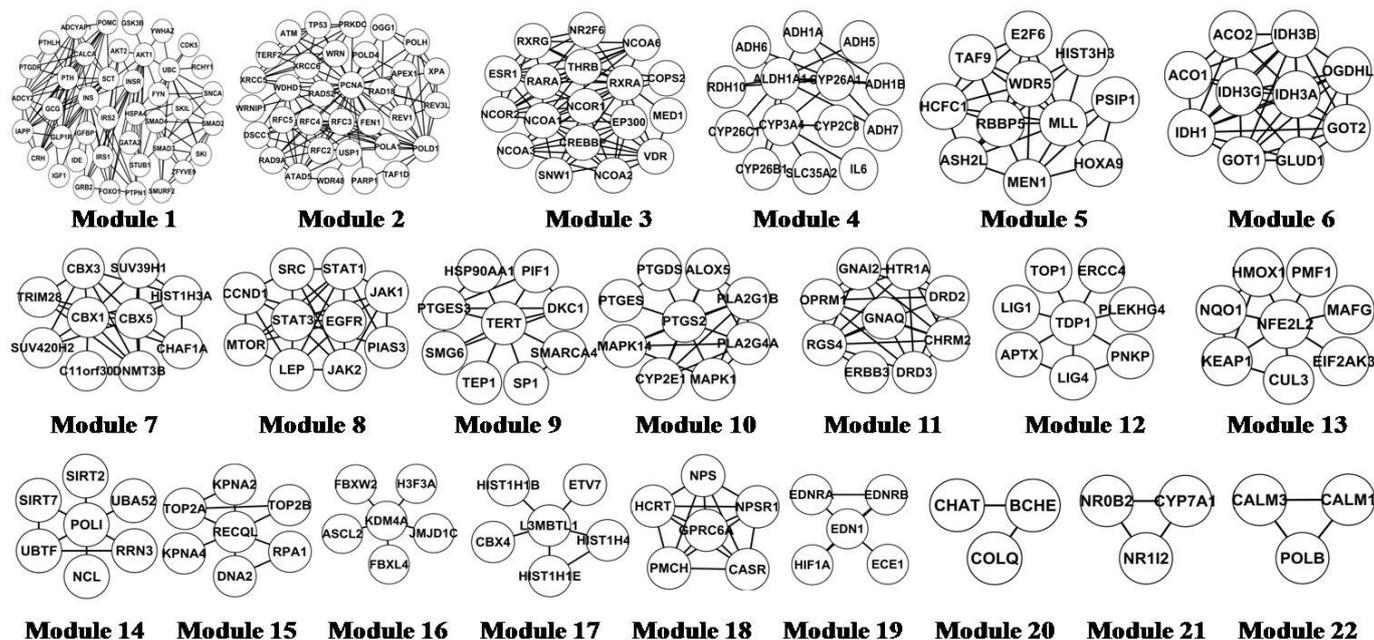


Fig. 3. Modules in the PIN of CPT. With the MCL algorithm, 22 modules are extracted from the network.

suppression of general protein translation [40]. Kinoshita and Yabuta showed that EIF2AK3 could induce cell apoptosis via the ubiquitin-proteasome system [40]. It had been reported that CPT induced ER stress-mediated apoptosis [41]. This indicated that CPT may exert antitumor properties through activation of EIF2AK3.

C. Conclusion

In this paper, the PIN of CPT exhibited the properties of scale-free property, small world property and modular architecture based on analysis of topological parameters. A module-based network analysis approach was proposed to expound the anti-inflammatory mechanism of CPT. The antitumor effects of CPT may be partly attributed to inhibiting the activation of the c-Src pathway and overexpression of EGFR, to mediating overexpression of PIAS and activation of EIF2AK3. Further experiments are needed to confirm the conclusions.

ACKNOWLEDGMENT

This work is financially supported by the National Key Technology R&D Program (No.2008BAI51B01) and the National natural fund project (No.81173522) in Beijing University of Chinese Medicine.

REFERENCES

[1] Zhang F, Zheng W, Pi R, Mei Z, Bao Y, Gao J, Tang W, Chen S and Liu P, "Cryptotanshinone protects primary rat cortical neurons from glutamate-induced neurotoxicity via the activation of the phosphatidylinositol 3-kinase/Akt signaling pathway," *Exp Brain Res*, vol. 193, pp. 109–118, February 2009.

[2] Zhou L, Zuo Z and Chow MS, "Danshen: an overview of its chemistry, pharmacology, pharmacokinetics, and clinical use," *J Clin Pharmacol*, vol. 45, pp. 1345–1359, December 2005.

[3] Wang BE, "Treatment of chronic liver diseases with traditional Chinese medicine," *J Gastroenterol Hepatol*, vol. 15 Suppl, pp. E67–E70, May 2000.

[4] Yu XY, Lin SG, Chen X, Zhou ZW, Liang J, Duan W, Chowbay B, Wen JY, Chan E, Cao J, Li CG and Zhou SF, "Transport of cryptotanshinone, a major active triterpenoid in *Salvia miltiorrhiza* Bunge widely used in the treatment of stroke and Alzheimer's disease, across the blood-brain barrier," *Curr Drug Metab*, vol. 8, pp. 365–378, May 2007.

[5] Lei Chen, Shi-zhong Zheng, Zhi-guang Sun, Ai-yun Wang, Chen-hu Huang, Neville A. Punchard, Shi-le Huang, Xiang Gao and Yin Lu, "Cryptotanshinone has diverse effects on cell cycle events in melanoma cell lines with different metastatic capacity," *Cancer Chemother Pharmacol*, vol. 68, pp. 17–27, July 2011.

[6] Kang BY, Chung SW, Kim SH, Ryu SY and Kim TS, "Inhibition of interleukin-12 and interferon-gamma production in immune cells by tanshinones from *Salvia miltiorrhiza*," *Immunopharmacology*, vol. 49, pp. 355–361, September 2000.

[7] Lee DS, Lee SH, Noh JG and Hong SD, "Antibacterial activities of cryptotanshinone and dihydrotanshinone I from a medicinal herb, *Salvia miltiorrhiza* Bunge," *Biosci Biotechnol Biochem*, vol. 63, pp. 2236–2239, December 1999.

[8] Lee HJ, Jung DB, Sohn EJ, Kim HH, Park MN, Lew JH, Lee SG, Kim B and Kim SH, "Inhibition of hypoxia inducible factor alpha and astrocyte-elevated gene-1 mediates cryptotanshinone exerted antitumor activity in hypoxic PC-3 cells," *Evid Based Complement Alternat Med*, vol. 2012, pp. 390957, November 2012.

[9] Li S, Zhang B. "Traditional Chinese medicine network pharmacology: theory, methodology and application," *Chin J Nat Med*, vol. 11, pp. 110–120, March 2013.

[10] Ivanov A.A., Khuri F.R. and Fu H., "Targeting protein-protein interactions as an anticancer strategy," *Trends in Pharmacological Sciences*, vol. 34, pp. 393–400, July 2013.

- [11] D. Pal, "On gene ontology and function annotation," *Bioinformatics*, vol. 1, pp. 97-98, February 2006.
- [12] Bento AP, Gaulton A, Hersey A, Bellis LJ, Chambers J, Davies M, Krüger FA, Light Y, Mak L, McGlinchey S, Nowotka M, Papadatos G, Santos R and Overington JP, "The ChEMBL bioactivity database: an update," *Nucleic Acids Res.*, vol. 42, pp. D1083-1090, January 2014.
- [13] Michael Kuhn, Damian Szklarczyk, Sune Pleischer-Frankild, Thomas H. Blicher, Christian von Mering, Lars J. Jensen, and Peer Bork, "STITCH 4: integration of protein-chemical interactions with user data," *Nucleic Acids Research*, vol. 42, pp. D401-D407, January 2014.
- [14] Franceschini A, Szklarczyk D, Frankild S, Kuhn M, Simonovic M, Roth A, Lin J, Minguez P, Bork P, von Mering C and Jensen LJ, "STRING v9.1: protein-protein interaction networks, with increased coverage and integration," *Nucleic Acids Res.*, vol. 41, pp. D808-815, January 2013.
- [15] Y Assenov, F Ramirez, SE Schelhorn, T Lengauer and M Albrecht, "Computing topological parameters of biological networks," *Bioinformatics*, vol. 24, pp. 282-284, January 2008.
- [16] Pržulj N, "Biological network comparison using graphlet degree distribution," *Bioinformatics*, vol. 19, pp. e177-e183, January 2007.
- [17] Enright AJ, Dongen SV and Ouzounis CA, "An efficient algorithm for large-scale detection of protein families," *Nucleic Acids Res.*, vol. 30, pp. 1575-1584, April 2002.
- [18] Sylvain Brohée and Jacques van Helden, "Evaluation of clustering algorithms for protein-protein interaction networks," *BMC Bioinformatics*, vol. 7, pp. 488, November 2006.
- [19] S Maere, K Heymans and M. Kuiper, "BiNGO: a Cytoscape plugin to assess overrepresentation of gene ontology categories in biological networks," *Bioinformatics*, vol. 21, pp. 3448-3449, November 2005.
- [20] Aggarwal BB, Sethi G, Ahn KS, Sandur SK, Pandey MK, Kunnumakkara AB, Sung B and Ichikawa H, "Targeting signal-transducer-and-activator-of-transcription-3 for prevention and therapy of cancer: modern target but ancient solution," *Ann N Y Acad Sci*, vol. 1091, pp. 151-169, December 2006.
- [21] Shin DS, Kim HN, Shin KD, Yoon YJ, Kim SJ, Han DC and Kwon BM, "Cryptotanshinone inhibits constitutive signal transducer and activator of transcription 3 function through blocking the dimerization in DU145 prostate cancer cells," *Cancer Res.*, vol. 69, pp. 193-202, January 2009.
- [22] Shannon P, Markiel A and Ozier O, "Cytoscape: a software environment for integrated models of biomolecular interaction networks," *Genome Res.*, vol. 13, pp. 2498-2504, November 2003.
- [23] Assenov Y, Ramirez F and Schelhorn S E, "Computing topological parameters of biological networks," *Bioinformatics*, vol. 24, pp. 282-284, January 2008.
- [24] Jeong H, Mason SP, Barabási AL and Oltvai ZN, "Lethality and centrality in protein networks," *Nature*, vol. 411, pp. 41-42, May 2001.
- [25] Alon U, Surette MG, Barkai N and Leibler S, "Robustness in bacterial chemotaxis," vol. 397, pp. 168-171, January 1999.
- [26] Callaway, Duncan S, MEJ Newman, SH. Strogatz and DJ Watts, "Network robustness and fragility: percolation on random graphs," vol. 85, pp. 5468-5471, December 2000.
- [27] SH Strogatz, "Exploring complex networks," *Nature*, vol. 410, pp. 268-276, March 2001.
- [28] Kisseleva T, Bhattacharya S, Braunstein, J and Schindler CW, "Signaling through the JAK/STAT pathway, recent advances and future challenges," *Gene*, vol. 285, pp. 1-24, February 2002.
- [29] Aaronson DS and Horvath CM, "A road map for those who don't know JAK-STAT," *Science*, vol. 296, pp. 1653-1655, May 2002.
- [30] Zhou JI, Xu XZ, Hu YR, Hu AR, Zhu CL and Gao GS, "Cryptotanshinone induces inhibition of breast tumor growth by cytotoxic CD4+ T cells through the JAK2/STAT4/ perforin pathway," *Asian Pac J Cancer Prev*, vol. 15, pp. 2439-2445, 2014.
- [31] Ogata Y, Osaki T, Naka T, Iwahori K, Furukawa M, Nagatomo I, Kijima T, Kumagai T, Yoshida M, Tachibana I and Kawase I, "Overexpression of PIAS3 suppresses cell growth and restores the drug sensitivity of human lung cancer cells in association with PI3-K/Akt inactivation." *Neoplasia (New York, N.Y.)*, vol. 8, pp. 817-825, October 2006.
- [32] Yu CL, Meyer DJ, Campbell GS, Lerner AC, Carter-Su C, Schwartz J and Jove R, "Enhanced DNA-binding activity of a Stat3-related protein in cells transformed by the Src oncoprotein," *Science*, vol. 269, pp. 81-83, July 1995.
- [33] Zhang Y, Turkson J, Carter-Su C, Smithgall T, Levitzki A, Kraker A, Krolewski JJ, Medveczky P and Jove R, "Activation of Stat3 in v-Src-transformed fibroblasts requires cooperation of Jak1 kinase activity," *J Biol Chem*, vol. 275, pp. 24935-24944, August 2000.
- [34] Garcia R, Bowman TL, Niu G, Yu H, Minton S, Muro-Cacho CA, Cox CE, Falcone R, Fairclough R, Parsons S, Laudano A, Gazit A, Levitzki A, Kraker A and Jove R, "Constitutive activation of Stat3 by the Src and JAK tyrosine kinases participates in growth regulation of human breast carcinoma cells," *Oncogene*, vol. 20, pp. 2499-2513, May 2001.
- [35] Dehm SM and Bonham K, "SRC gene expression in human cancer: the role of transcriptional activation." *Biochem. Cell Biol.*, vol. 82, pp. 263-274, April 2004.
- [36] Biscardi JS, Belsches AP and Parsons SJ, "Characterization of human epidermal growth factor receptor and c-Src interactions in human breast tumor cells," *Mol. Carcinog*, vol. 21, pp. 261-272, April 1998.
- [37] Walker F, Abramowitz L, Benabderrahmane D, Duval X, Descatoire V, Hélin D, Lehy T and Aparicio T. "Growth factor receptor expression in anal squamous lesions: modifications associated with oncogenic human papillomavirus and human immunodeficiency virus," *Hum. Pathol*, vol. 40, pp. 1517-1527, November 2009.
- [38] Araujo JA, Zhang M and Yin F, "Heme oxygenase-1, oxidation, inflammation, and atherosclerosis," *Front Pharmacol*, vol. 3, pp. 119, July 2012.
- [39] Oommen D and Prise KM, "Down-regulation of PERK enhances resistance to ionizing radiation," *Biochem Biophys Res Commun*, vol. 441, pp. 31-35, November 2013.
- [40] Yahiro K, Tsutsuki H, Ogura K, Nagasawa S, Moss J and Noda M, "Regulation of subtilase cytotoxin-induced cell death by an RNA-dependent protein kinase-like endoplasmic reticulum kinase-dependent proteasome pathway in HeLa cells," *Infect Immun*, vol. 80, pp. 1803-1814, May 2012.
- [41] Park II, Kim MJ, Park OJ, Choe W, Kang I, Kim SS and Ha J, "Cryptotanshinone induces ER stress-mediated apoptosis in HepG2 and MCF7 cells," *Apoptosis*, vol. 17, pp. 248-257, March 2012.